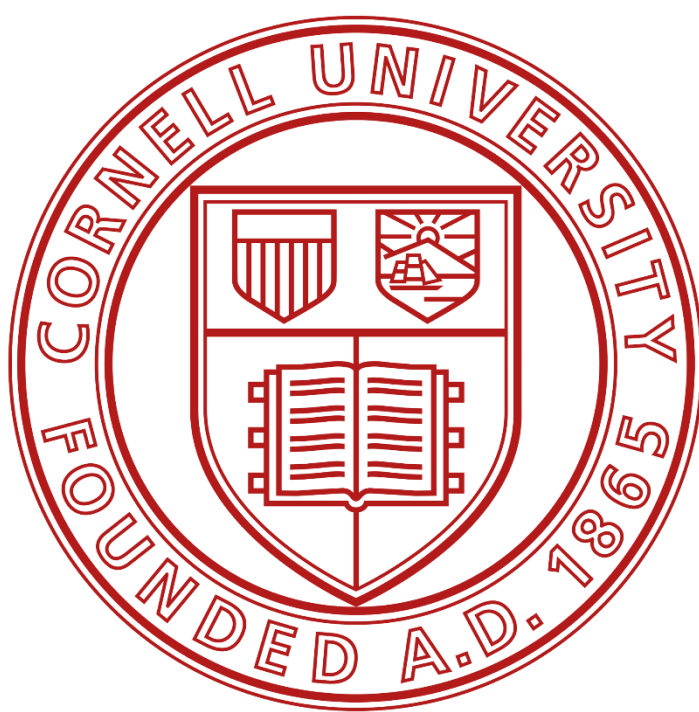
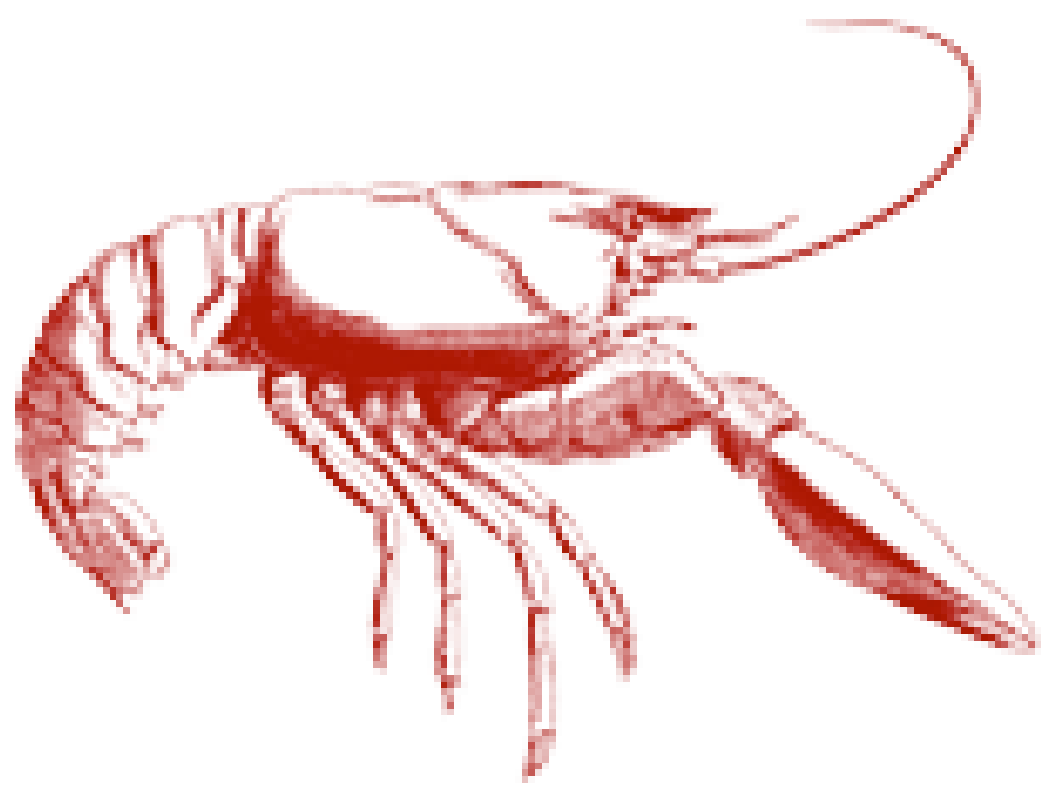




Low-Cost Extracellular Voltage Amplifier for Neural Signal Acquisition



Author: Gilbert Liang **Advisors:** Bruce R. Land, Van Hunter Adams, Bruce R. Johnson

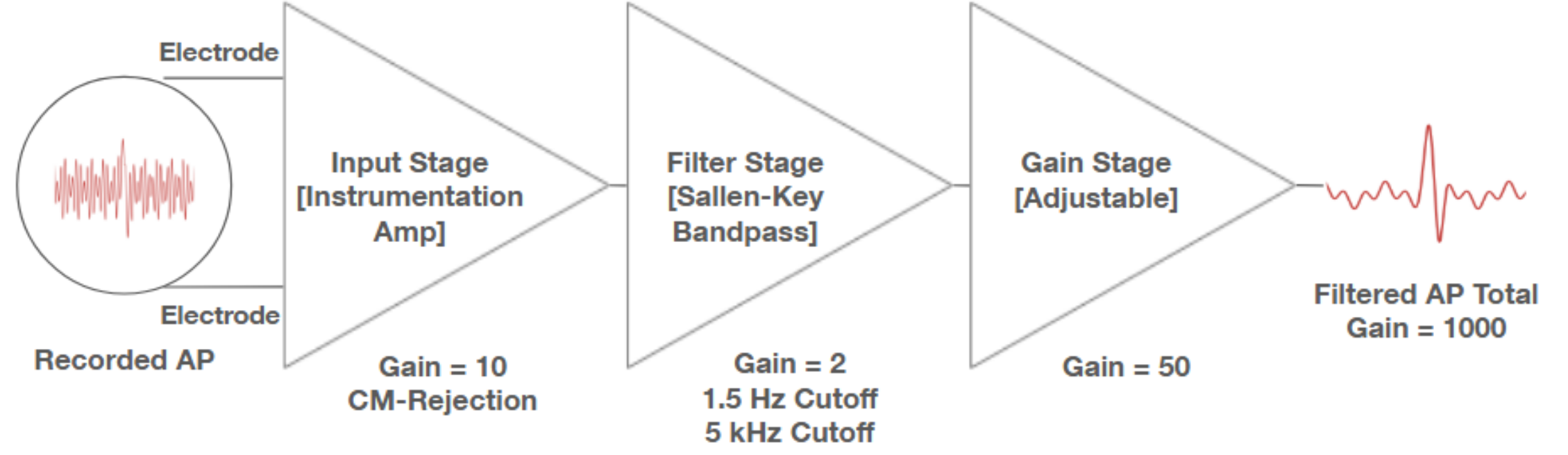
Cost Barriers in Neurobiological Experiments

Extracellular voltage-amplifiers are necessary for measuring action potential (AP) signals in neurobiological experiments. Commercial amplifiers can cost from \$500 to \$1000 per device-channel. By designing a **low-cost extracellular voltage amplifier** that has comparable functionality to the commercial devices available this provides additional access to studying neural signals in both educational and *limited* resource research settings.

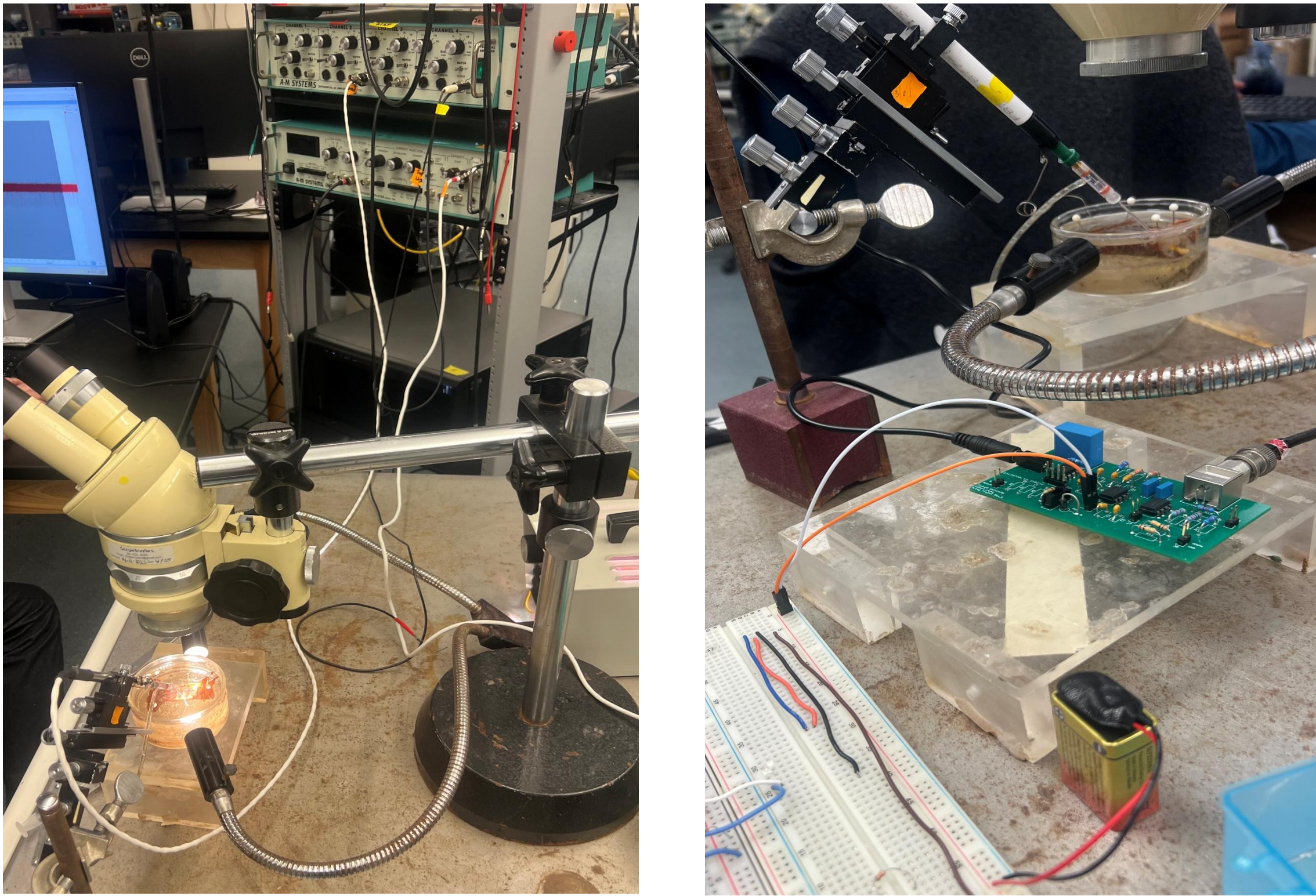
Amplifier Design Goals

- Designing Based on A-M 1700 Differential AC Amplifier:
1. Simple Enough Design for DIY (Low Cost)
 2. Gain (Adjustable): 100 & 1000
 3. Frequency Response: 1.5 Hz to 5 kHz
 4. High Common-Mode-Rejection-Ratio (CMRR)
 5. High Signal-to-Noise Ratio (SNR)
 6. High Input Impedance
 7. Powered using 9V-battery → Minimal power consumption
 8. Minimal Sizing and Area

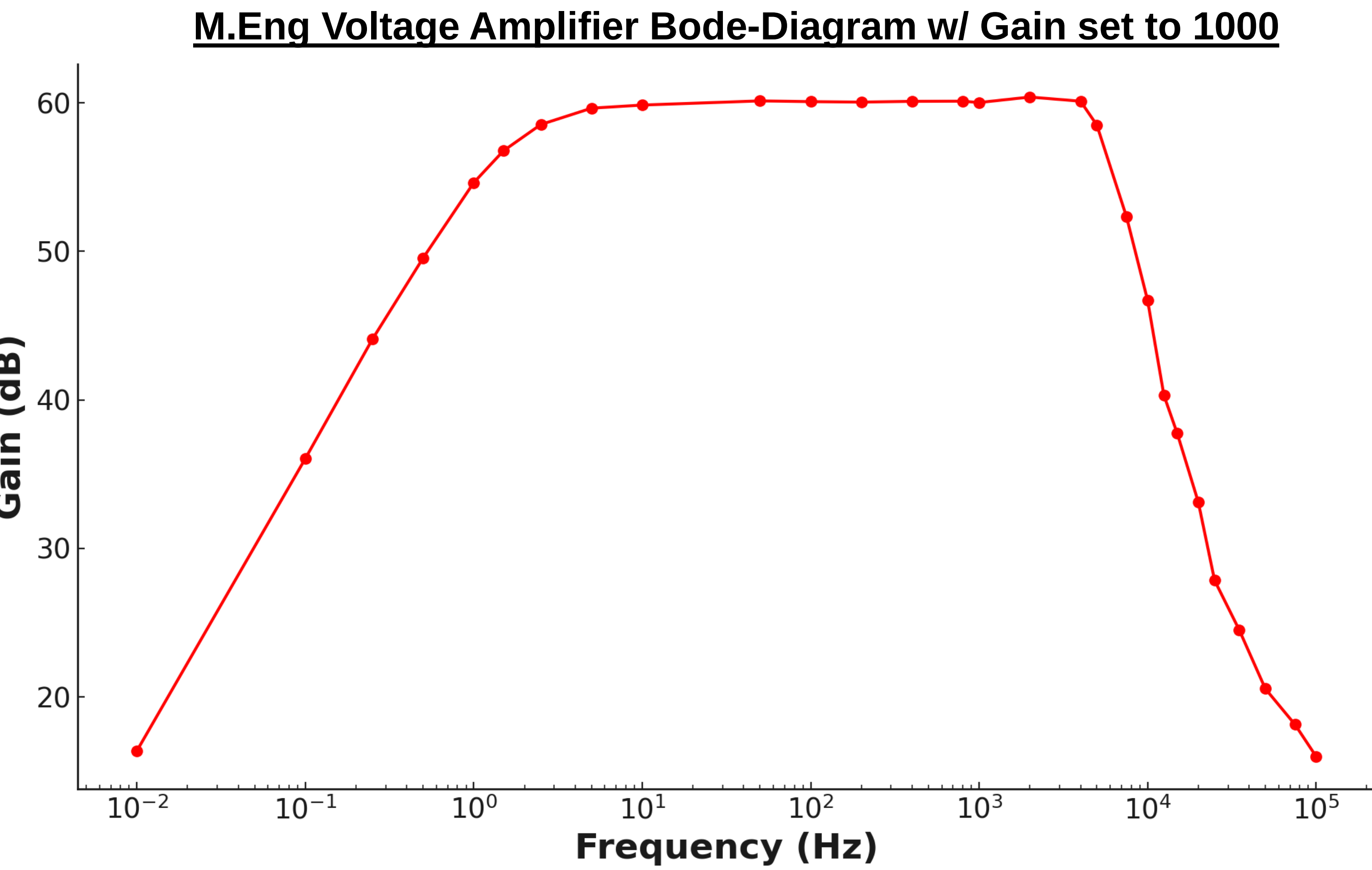
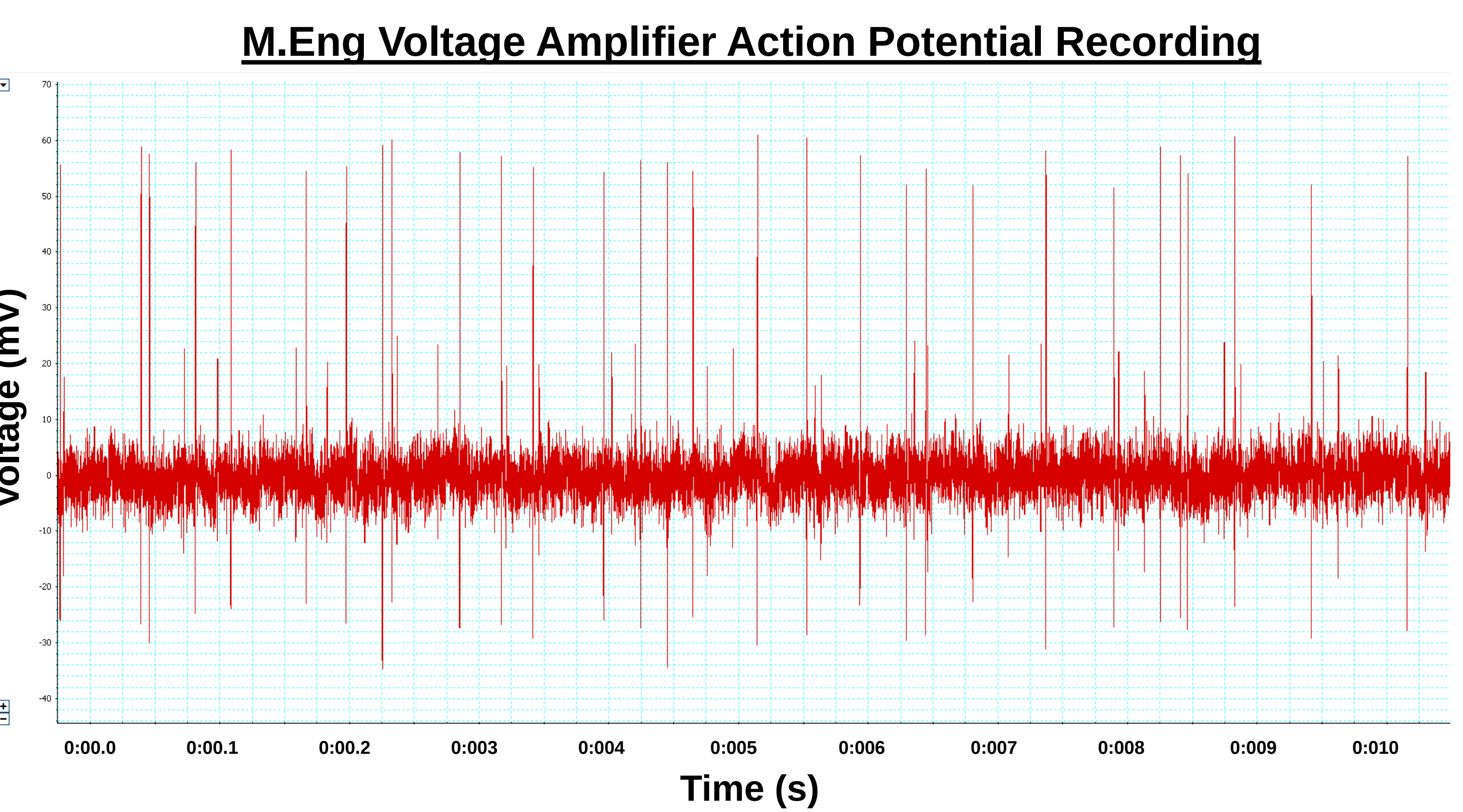
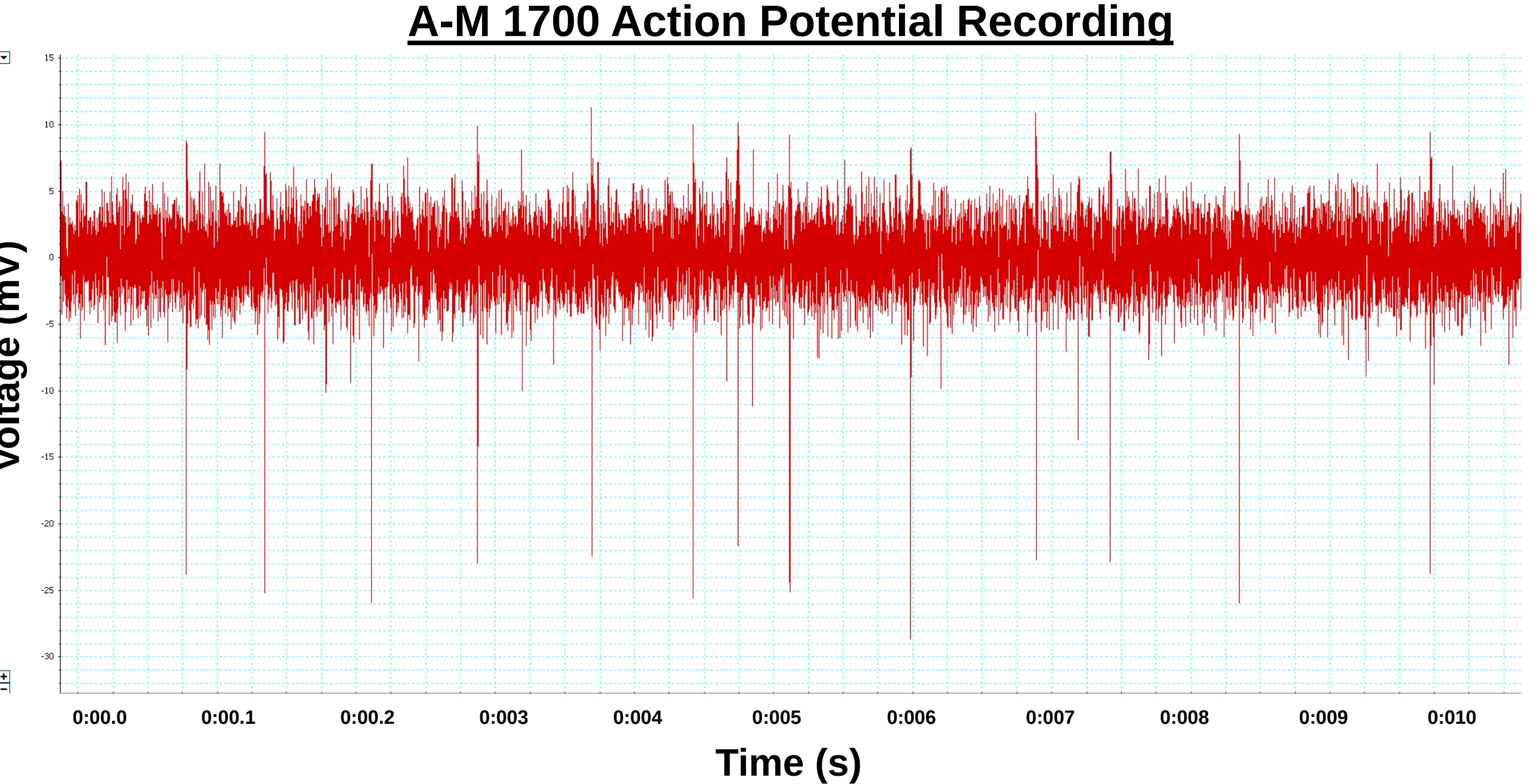
Device Block Diagram



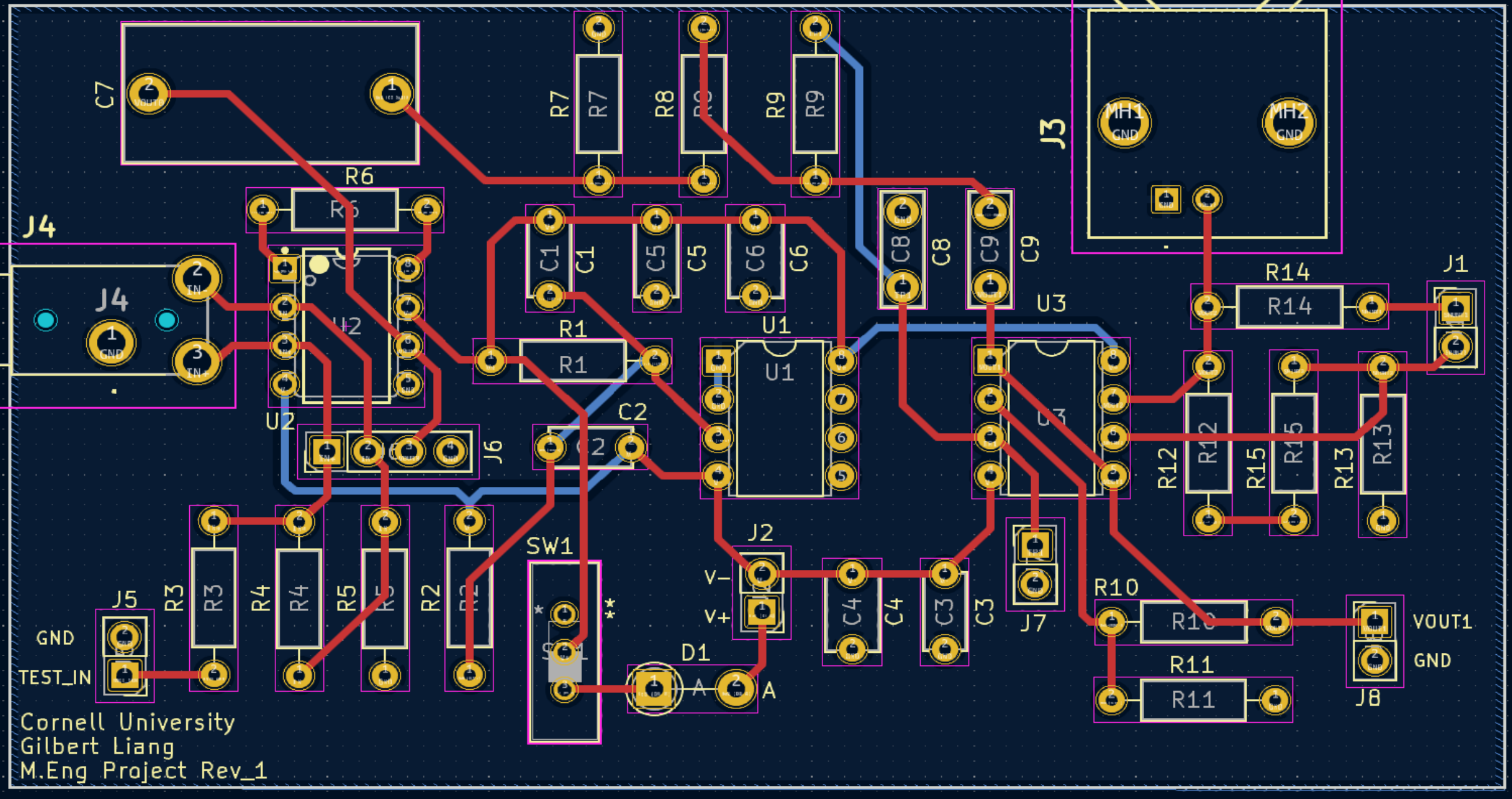
Crawdad Lab Experiment Setup



Crawdad Motor Nerve Recording: Comparable to Commercial Amplifier



Amplifier PCB Layout



Size: 92.4mm x 72.21mm x 19.56mm

Comparing Device Functionality

	A-M 1700 Spec.	M.Eng PCB Spec.
Gain (Adjustable):	100 1000 10,000*	100 1000
Frequency Response:	300Hz - 5kHz*	1.5Hz - 5kHz
Signal-to-Noise Ratio (SNR)	~ 3.5:1 (avg)	~ 4.25:1 (avg)
Input Impedance	100G Ω	1T Ω
Power Consumption	-	1.8mA @ 9V (16.2mW)
Cost	\$3130 (4-Channels)	\$32 (1-Channel)

Conclusion & Next Steps

A functional extracellular voltage amplifier was designed and tested in a lab experiment, recording the action potential signals from a stimulated motor nerve. When compared to the \$782.5 (per device-channel) A-M 1700, the **\$32 single-channel device** can provide the same gain at the defined cut-off frequencies with similar SNR while powered by a 9V battery.

Next steps:

1. Design metal-enclosure for PCB-assembly
2. Increase low cut-off frequency to 300Hz to stabilize noise floor recording
3. Add adjustable frequency functionality and multi-channel output

References

[1] B. R. Land, R. A. Wytenbach, and B. R. Johnson, "Tools for teaching neurophysiology: The Backyard Brains SpikerBox and other projects." *J. Neurosci. Methods*, vol. 106, no. 1, pp. 47–55, Jan. 2001, doi: 10.1016/S0165-0270(01)00341-6.

[2] B. R. Johnson, "Crawdad Lab 2: Motor nerve recording," *Emory University Crawdad Lab Manual*, [Online]. Available: <https://faculty.college.emory.edu/sites/wytenbach/crawdad/2.NerveRecording.html>.

[3] A-M Systems, "Model 1700 Differential AC Amplifier," *Biomedical Instruments Trading*, [Online]. Available: <https://www.biomedical-instruments-trading.com/products/electrophysiology/amplifiers/differential-extracellular/84-a-m-systems-model-1700>.

Acknowledgements

I would like to thank Professor Bruce Land and Professor Hunter Adams for their guidance and unwavering support throughout the project. I would also like to thank Pawan Perera (Cornell ECE, Class of SP2024) for the initial circuit schematic design of this device. Additionally, would like to thank Cornell University and the M.Eng Program for sponsoring this project.

I would also like to thank Professor Bruce Johnson and the BioNB/BME/ECE 4910 Student Neurophysiology lab for providing the lab test setup, equipment and the procedures to perform "Crawdad" to verify this project.

